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## A SPIROSTANOL GLYCOSIDE FROM *AGAVE CANTALA*

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**Key Word Index**—*Agave cantala*; Agavaceae; rhizomes; saponin; spirostanol glycoside; tigogenin;  $^{13}\text{C}$ INEPT and  $^1\text{H}$  decoupled NMR.

**Abstract**—A new spirostanol glycoside, cantalasaponin-3, isolated from the methanolic extract of the rhizomes of *Agave cantala*, has been characterized.

### INTRODUCTION

*Agave* species have been used for medicinal purposes and various saponins have been reported from *A. cantala* Roxb. [1]. This communication deals with the structure elucidation of cantalasaponin-3 (2) isolated from the rhizomes of this plant.

### RESULTS AND DISCUSSION

Saponin 2, a 25R-spirostan derivative (IR) was found to have an  $M_r$  of 1034, as indicated from the pseudo-molecular ions at  $m/z$  1073, 1057 and 1035 corresponding to  $[\text{M} + \text{K}]^+$ ,  $[\text{M} + \text{Na}]^+$  and  $[\text{M} + \text{H}]^+$  ions, respectively, in its FD-mass spectrum. The peaks at  $m/z$  925/903 and at 895/873 arise from the loss of terminal pentose and hexose, respectively, from  $[\text{M} + \text{Na}]^+ / [\text{M} + \text{H}]^+$  ions.

Acidic hydrolysis of 2 gave tigogenin, and D-galactose, D-glucose and D-xylose in the ratio 1:2:1.

The interglycosidic linkages in 2 were established by means of  $^{13}\text{C}$  NMR spectroscopy.  $^{13}\text{C}$  chemical shifts of methyl pyranosides of  $\beta$ -D-galactose,  $\beta$ -D-glucose and  $\beta$ -D-xylose in pyridine- $d_5$  [2–4] and those of tigogenin [5] are available and the signals in 2 were assigned by the application of glycosylation shifts [2, 3]. In the  $^{13}\text{C}$ INEPT spectrum, by setting the delay time  $\Delta$  as  $3/4J$  [6], CH and Me signals were in phase,  $\text{CH}_2$  out of phase, and quaternary carbons and carbons of the solvent were absent. In the  $^1\text{H}$  decoupled mode the signals in the sugar region of 2 and 1 [1], the 12-oxo analogue of 2, were almost superimposable. This observation was further supported when the permethylation products of 2 and its partial hydrolysis product,  $\text{PS}_2$ , gave methylated sugars identical to those obtained after permethylation of 1 and  $\text{PS}_3$  [1], respectively.

The anomeric linkages were deduced as  $\beta$  from the  $^1\text{H}$  NMR spectrum of 2 and by the application of Klyne's rule [7].

Thus, 2 was characterized as 3-O- $\{\beta$ -D-glucopyranosyl(1  $\rightarrow$  3)- $\beta$ -D-glucopyranosyl(1  $\rightarrow$  2) $\}$   $\{\beta$ -D-xylopyranosyl(1  $\rightarrow$  4)- $\beta$ -D-galactopyranosyl]- (25R)-5 $\alpha$ -spirostan-3 $\beta$ -ol, a 12-deoxo analogue of 1 [1]. This provides an example of the co-occurrence of hecogenin and tigogenin glycosides with identical sugar chains.

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## EXPERIMENTAL

Almost all the instrumentation techniques were as described in ref. [1].  $^{13}\text{C}$ INEPT and  $^1\text{H}$  decoupled,  $^1\text{H}$ NMR spectra were recorded on a JEOL FX-100 Fourier-transform spectrometer operating at 25/100 MHz. Isolation of **2** and the solvent systems used for the TLC examination of sugars and methylated sugars are detailed in ref. [1]. The compounds on TLC were visualized with 10% ethanolic  $\text{H}_2\text{SO}_4$  and on prep. TLC by spraying with  $\text{H}_2\text{O}$ .

**Compound 2.** Colourless plates (130 mg) from aq. EtOH, mp 298–302°,  $[\alpha]_{\text{D}}^{25-17} -54.8^\circ$  ( $\text{C}_5\text{H}_5\text{N}$ ; c 1.29). IR  $\nu_{\text{max}}^{\text{KBr}}$   $\text{cm}^{-1}$ : 3400 (OH), 985, 925, 900, 870 (intensity 900 > 925, 25R-spiroketal); FDMS,  $m/z$  (rel. int.): 1073  $[\text{M} + \text{K}]^+$  (10.1), 1057  $[\text{M} + \text{Na}]^+$  (100.0), 1035  $[\text{M} + \text{H}]^+$  (11.9), 925  $[\text{M} + \text{Na} - 132]^+$  (27.8), 903  $[\text{M} + \text{H} - 132]^+$  (23.0), 895  $[\text{M} + \text{Na} - 162]^+$  (10.5), 873  $[\text{M} + \text{H} - 162]^+$  (10.5), 741  $[\text{M} + \text{H} - 132 - 162]^+$  (3.8), 579  $[\text{M} + \text{H} - 132 - 162 - 162]^+$  (4.3), 399  $[\text{genin} + \text{H} - \text{H}_2\text{O}]^+$  (7.2), 163  $[\text{hex} + \text{H} - \text{H}_2\text{O}]^+$  (8.3), 133  $[\text{xyz} + \text{H} - \text{H}_2\text{O}]^+$  (14.0);  $^1\text{H}$ NMR:  $\delta$  0.67 (6H, m, 18-Me, 27-Me), 0.82 (3H, s, 19-Me), 1.12 (3H, d,  $J = 7$  Hz, 21-Me), 4.85 (1H, d,  $J = 7.1$  Hz, H-1 of glu), 5.20 (1H, d,  $J = 7.3$  Hz, H-1 of glu), 5.26 (1H, d,  $J = 7$  Hz, H-1 of gal), 5.61 (1H, br s, H-1 of xyz);  $^{13}\text{C}$ NMR: aglycone:  $\delta$  37.2, 29.9, 78.6, 34.8, 44.6, 29.0, 32.4, 35.3, 54.4, 35.8, 21.3, 40.1, 40.8, 56.4, 31.8, 81.1, 63.0, 16.6, 12.3, 42.0, 15.0, 109.2, 32.1, 28.9, 30.6, 66.9, 17.3 (C-1–C-27); sugar moiety:  $\delta$  102.4, 81.1, 73.1, 79.9, 76.2, 60.2 (galactosyl C-1–C-6), 104.9, 70.7, 86.8, 70.4, 77.6, 62.4<sup>a</sup> (glucosyl C-1–C-6), 104.9, 75.5, 78.6, 71.0, 78.6, 63.0<sup>a</sup> (glucosyl C-1'–C-6'), 104.9, 75.1, 77.4, 70.7, 67.3 (xylosyl C-1–C-5). (Found: C, 58.19; H, 7.99.  $\text{C}_{50}\text{H}_{82}\text{O}_{22}$  requires C, 58.03; H, 7.93%.)

**Acidic hydrolysis of 2.** Compound **2** (15 mg) was refluxed with 2 M HCl–MeOH (1:1, 8 ml) on a boiling water bath for 3 hr to afford the aglycone (tigogenin), mp 203–205°,  $[\alpha]_{\text{D}}^{20} -64^\circ$  ( $\text{CHCl}_3$ ; c 1.0). IR  $\nu_{\text{max}}^{\text{KBr}}$   $\text{cm}^{-1}$ : 3400 (OH), 980, 920, 900, 864 (intensity 900 > 920); EIMS:  $m/z$  416  $[\text{M}]^+$ . The neutralized and conc. aq. hydrolysate contained D-galactose, D-glucose and

D-xylose (PC). Sugars were estimated by colorimetry using a wavelength of 420 nm.

**Partial hydrolysis of 2.** Compound **2** (70 mg) in 1 M HCl–*n*-BuOH (1:1, 20 ml) was heated at 70° for 2 hr. The BuOH layer was washed with 5%  $\text{NaHCO}_3$  and then with  $\text{H}_2\text{O}$  and concd *in vacuo* to afford a residue, which was purified by prep. TLC ( $\text{CHCl}_3$ –MeOH– $\text{H}_2\text{O}$ , 13:4:2) to give tigogenin (2.5 mg),  $\text{PS}_1$  (6 mg) and  $\text{PS}_2$  (20 mg).

$\text{PS}_1$ . Colourless plates from MeOH, mp 232–235°,  $[\alpha]_{\text{D}}^{20} -38.0^\circ$  ( $\text{C}_5\text{H}_5\text{N}$ ; c 1.0).  $\text{PS}_1$  (4 mg) on hydrolysis gave D-galactose and was found to be identical with tigogenin-D-galactoside [8] (IR).

**Permethylation of 2 and  $\text{PS}_2$ .** Compounds **2** and  $\text{PS}_2$  (14 mg each) were separately permethylated with MeI (3 ml) and  $\text{Ag}_2\text{O}$  (200 mg) in DMF (0.5 ml). Usual work-up gave syrups which were purified by prep. TLC ( $\text{C}_6\text{H}_6$ – $\text{Me}_2\text{CO}$ , 4:1) to yield **2a** (7 mg) and  $\text{PS}_{2a}$  (6 mg).

**Hydrolysis of 2a and  $\text{PS}_{2a}$ .** Compound **2a** and  $\text{PS}_{2a}$  (6 mg each) were separately refluxed with 1 M HCl–MeOH (1:1, 5 ml) for 3.5 hr and 2 hr, respectively. The neutralized and concd hydrolysate from **2a** contained 2,3,4,6-tetra-*O*-methyl-D-glucose, 2,3,4-tri-*O*-methyl-D-xylose, 2,4,6-tri-*O*-methyl-D-glucose and Wallenfels' positive 3,6-di-*O*-methyl-D-galactose, all identical to those in the hydrolysate of **1a**.  $\text{PS}_{2a}$  gave methylated sugars identical to those in the hydrolysate of the permethylate of  $\text{PS}_3$  [1].

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<sup>13</sup>CNMR data: <sup>a</sup>Assignments are interchangeable between carbons marked with similar sign.